

TriLink
BioTechnologies

GATCAGTACAAAGC
TTTCTGAGTACAAAGC
TATTTAGAAGTACAAAGC
ACAGAAAGTACAAAGC
AAATAGAAAGTACAAAGC
CTGGAAAGTACAAAGC
TTTGAAGTACAAAGC
TTTGAAGTACAAAGC
CAGAAAGTACAAAGC
GTAGTGAATAAAAGC
ACTTFTACTAGAAAGC
ATAGAGATGTATCAAGC
CTTFTACTAGAAAGC
ATAGAGATGTATCAAGC
GGAAATACAAAGC
CAGAAAGTACAAAGC
AAGTACATTTGGAAAGC
GTGAATAAAAGC
TAGACATGTATCAAGC
GGAAATACAAAGC
TAGAACTTCGAAGC
AAGTACATTTGGAAAGC
GTGAATAAAAGC
AAGTACATTTGGAAAGC
TAGAGATGTATCAAGC
GGAAATACAAAGC
TAGAACTTCGAAGC
AAGTACATTTGGAAAGC
CTTFTACTAGAAAGC
ATAGAGATGTATCAAGC
GTGAATAAAAGC
TAGAGATGTATCAAGC
GGAAATACAAAGC

Enzymatic Incorporation of Biotin-16-AA-dNTPs

TriLink BioTechnologies, Inc.
Research and Development

Contributors: Joyclyn Yee, Stephanie Perry,
Michelle McNamara, Natasha Paul

Overview

Biotin is a naturally occurring cofactor that binds very tightly to the tetrameric protein, avidin. This strong association between biotin and avidin has been used in a number of biotechnology applications, which include detection and isolation of a molecule of interest. One approach for introducing biotin modifications into DNA is by the enzymatic incorporation of nucleoside 5'-triphosphates that have been chemically modified with biotin.

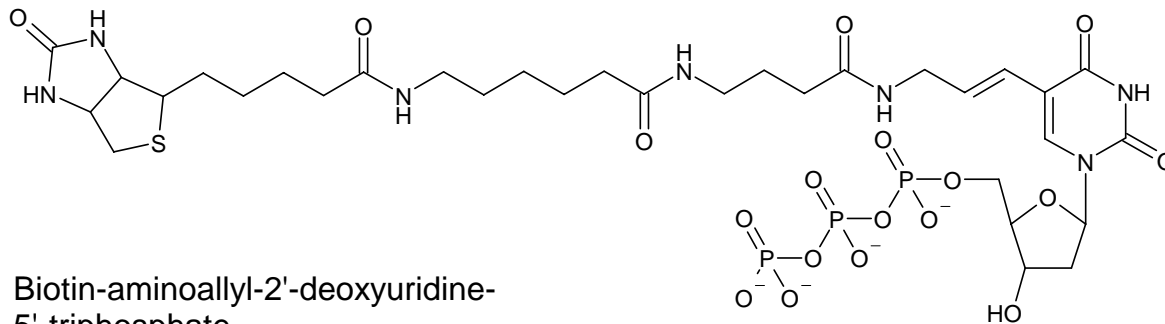
Outline

- Goal: Evaluate the enzymatic incorporation of Biotin-16-AA-dNTPs.

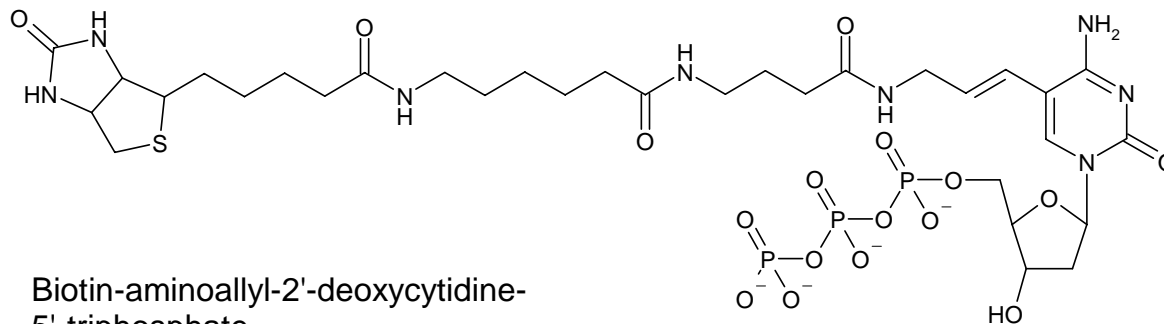
- Approach:

- Evaluate Biotin-16-AA-dNTP incorporation in primer extension experiments using either MMLV reverse transcriptase or Klenow(exo-) DNA polymerase.
- Evaluate the incorporation of Biotin-16-AA-dNTPs in PCR.

Chemical Structures of the Biotin-16-AA-dNTPs



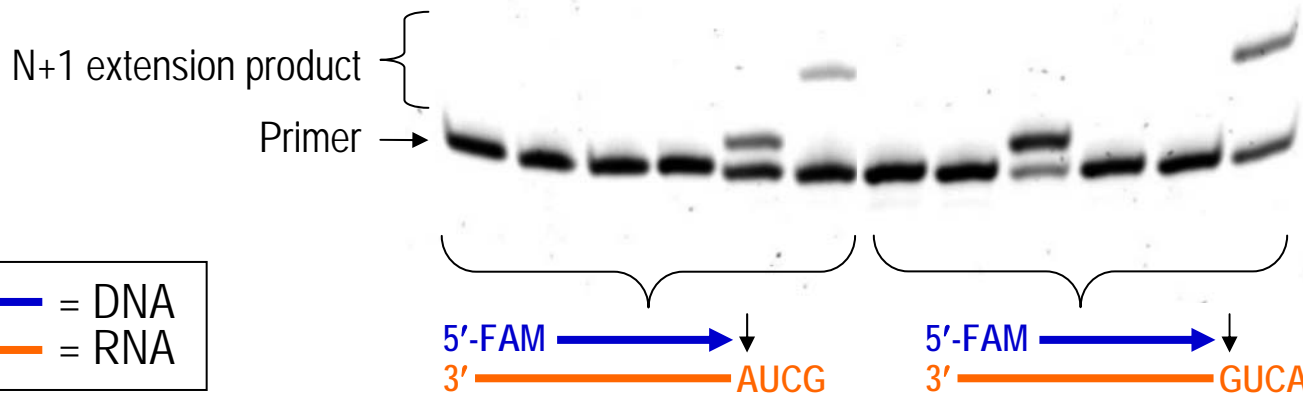
Biotin-aminoallyl-2'-deoxyuridine-5'-triphosphate



Biotin-aminoallyl-2'-deoxycytidine-5'-triphosphate

Single Nucleotide Incorporation of Biotin-16-AA-dNTPs by MMLV Reverse Transcriptase

| | | | | | | | | | | | |
|-------------------|--|---|---|---|---|---|---|---|---|---|---|
| dATP | | + | | | | | + | | | | |
| dCTP | | | + | | | | | + | | | |
| dGTP | | | | + | | | | | + | | |
| dTTP | | | | | + | | | | | + | |
| Biotin-16-AA-dCTP | | | | | | | | | | | + |
| Biotin-16-AA-dUTP | | | | | | + | | | | | |

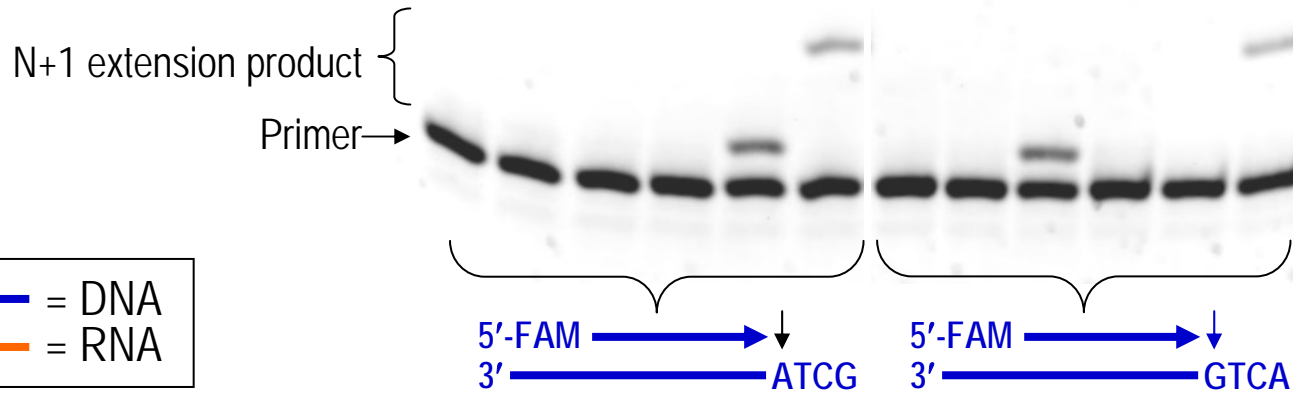


All biotinylated dNTPs are substrates for MMLV reverse transcriptase and produce an extension product with lower mobility.

Experimental Conditions: 1x First strand synthesis buffer (50 mM Tris-HCl (pH 8.3 @ 25°C), 50 mM KCl, 3.0 mM MgCl₂, 5 mM DTT), 5'-FAM-labeled primer (6.25 μM), RNA template (10 μM), Ambion MMLV Reverse transcriptase (0.2 U/μL), 0.1 mM dNTP. Thermal cycling parameters: 42°C @ 20 min; 98°C @ 2 min.

Single Nucleotide Incorporation of Biotin-16-AA-dNTPs by Klenow(exo-) DNA Polymerase

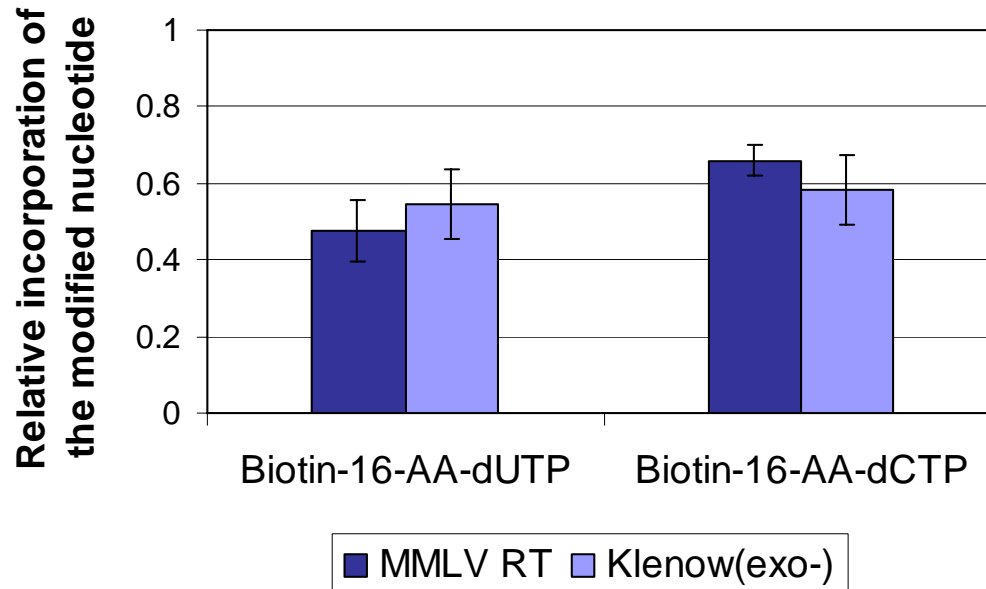
| | | | | | | | | | | | |
|-------------------|--|---|---|---|---|---|---|---|---|---|---|
| dATP | | + | | | | | + | | | | |
| dCTP | | | + | | | | | + | | | |
| dGTP | | | | + | | | | | + | | |
| dTTP | | | | | + | | | | | + | |
| Biotin-16-AA-dCTP | | | | | | | | | | | + |
| Biotin-16-AA-dUTP | | | | | | + | | | | | |



All biotinylated dNTPs are substrates for Klenow(exo-) DNA polymerase and produce an extension product with lower mobility.

Experimental Conditions: 1X NEBuffer 2 (10 mM Tris-HCl (pH 7.9 @ 25°C), 50 mM NaCl, 10 mM MgCl₂, 1 mM DTT), 5'-FAM-labeled primer (10 μM), DNA template (17 μM), New England Biolabs Klenow(exo-) DNA polymerase (0.025 U/μL), 20 μM dNTP. Thermal cycling parameters: 37°C @ 20 min; 72°C @ 20 min.

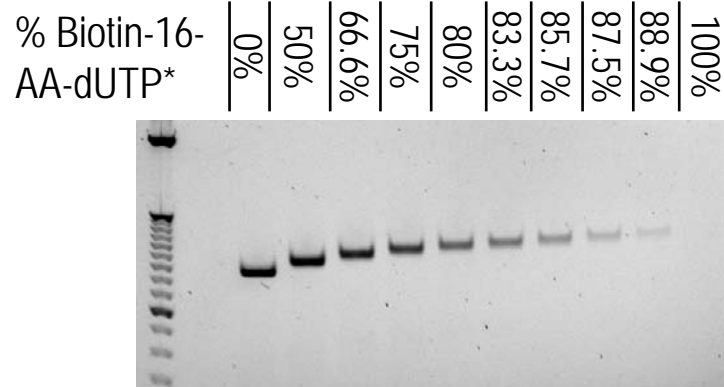
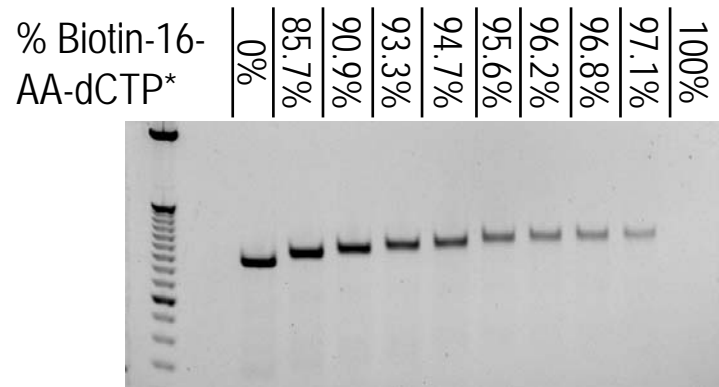
Summary of biotin-16-AA-dNTP Single Nucleotide Incorporation Studies



$$\text{Relative modified nucleotide incorporation} = \frac{\% \text{ n+1 extension product (modified dNTP)}}{\% \text{ n+1 extension product (natural dNTP)}}$$

Relative to their natural counterpart, biotinylated dCTP and dUTP analogs are incorporated with ~50% lower efficiency at the conditions examined.

Incorporation of Biotinylated dNTPs by *Taq* DNA Polymerase in PCR



*Total concentration of biotinylated + natural dNTP was maintained at 0.2 mM. The percentage of biotinylated dNTP was titrated between 0 and 100%.

*All biotinylated dNTPs are substrates for *Taq* DNA polymerase in PCR.*

As the percentage of biotinylated nucleotide increases, a corresponding decrease in the mobility of the amplicon is observed.

100% biotinylated dNTP substitution causes complete reaction inhibition.

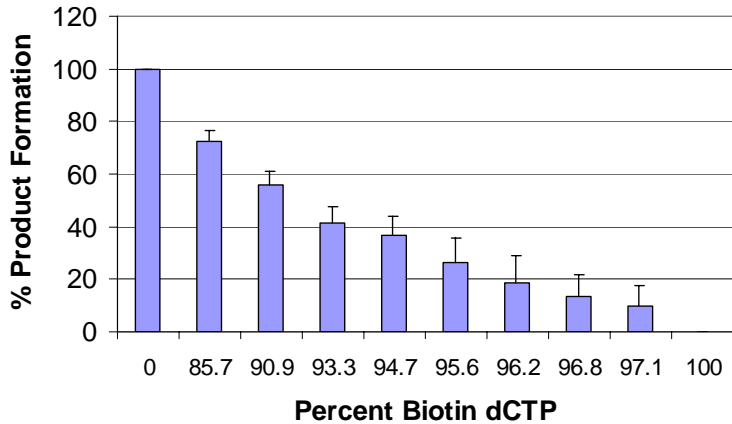
Experimental Conditions: Control Lambda primers (0.2 mM), New England Biolabs *Taq* DNA Polymerase (1 U/50 mL rxn), 0.2 mM dNTPs (including Biotin and natural), 1x NEB buffer (10 mM Tris-HCl (pH 8.3 @ 25°C), 50 mM KCl, 1.5 mM MgCl₂, Lambda genomic DNA (1 ng/50 µL rxn).

Primer sequences: 5'-CCTGCTTCTGCCGCTTCACGA and 5'-TCCGGATAAAAACGTCGATGACATTTGC.

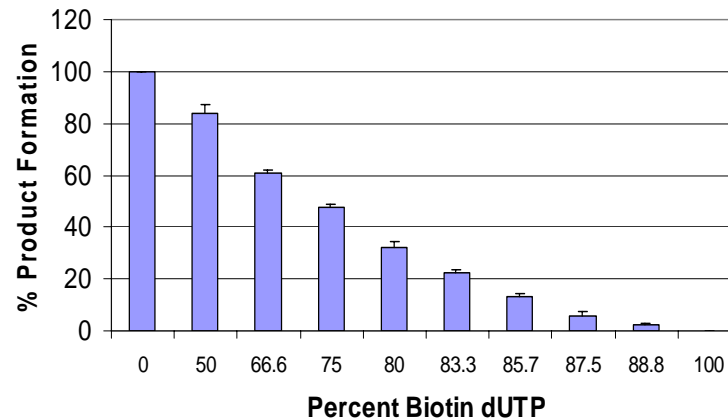
Thermal cycling parameters: 95°C @ 2 min; [95°C @ 15 sec, 55°C @ 15 sec, 72°C @ 45 sec]25x; 72°C @ 5 min.

Incorporation of Biotinylated dNTPs by *Taq* DNA Polymerase in PCR

Biotin-16-AA-dCTP



Biotin-16-AA-dUTP



The percent product formation is the product yield, normalized to the yield with 100% unmodified dNTPs.

Extent of biotinylated dNTP substitution that yields ~50% PCR product formation:

Biotin dCTP = ~92%

Biotin dUTP = ~75%

Summary

- *Biotin-16-AA-dCTP and biotin-16-AA-dUTP can be readily incorporated by reverse transcriptases and DNA polymerases in primer extension schemes.*
- *Biotinylated nucleotides are readily incorporated during PCR amplification schemes.*
- *Biotin-16-AA-dCTP can be substituted to a greater extent for its natural dNTP in PCR without compromising amplicon yield.*



TriLink
BioTechnologies

GAAGTACATTGGGAG
TTTCTGGGAG
TATTTAGA
ACAGAAAGT
AAATAGAG
CTGGAAATACAAAGT
TTTAGAACTTGGAA
CAGAAAGTACATTGGGAG
GTAGTGAATAAAAGCT
ACTTTTACTAGAAAG
ATAGAGATGTATCA
CTTTTACTAGAAAG
ATAGAGATGTATCA
TGGAAATACAAAGT
CAGAACTTGGGAG
AAGTACATTGGGAG
GTGAATAAAAGCT
TAGAGATGTATCA
GGAAATACAAAGT
TAGAACTTGGGAG
AAGTACATTGGGAG
GTGAATAAAAGCT
AAGTACATTGGGAG
TAGAGATGTATCA
GGAAATACAAAGT
TAGAACTTGGGAG
AAGTACATTGGGAG
CTTTTACTAGAAAG
ATAGAGATGTATCA
GTGAAT
TAGAGATGTATCA
CTGGG