Development of Standardized Pathogen Detection Assays Using CleanAmp™ Master Mixes

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Abstract

Acute gastroenteritis (AGE) affects over 179 million people and causes over 600,000 hospitalizations and 5000 deaths in the U.S. annually (CDC). Infectious AGE can be caused by a wide variety of different pathogens including bacteria, viruses and parasites. Rapid, definitive and economical identification of the causative AGE agent could inform medical decisions while reducing the inappropriate use of antibiotics. Standardized molecular methods and willing large and the ace identification area officiant and aceamplical and the standardized molecular methods. multiplexing will make pathogen identification more efficient and economical, with a huge potential global impact.

The molecular diagnostics field demands assays with more speed, specificity, and sensitivity. PCR is an established and reliable technology that meets these criteria. Although there are a myriad of published protocols using PCR for detection of specific diseases there is still a lack of established and standardized assays that cover the detection of certain infections.

CleanAmp[™] dNTPs are a universal Hol Start technology that can be applied to many different types of PCR assays including Real-Time PCR. Multiplex PCR and RTPCR. CleanAmp[™] dNTPs are 2' deaxynucleotide triphosphates with a 3' thermolabile protecting group that prevents incorporation of the dNTPs at room temperature. As temperature is increased the protecting group begins to fail off creating a standard dNTP that can be naturally incorporated into the growing cDNA or PCR strand. CleanAmp[™] dNTPs do not have a specific activation temperature but their kinetics are determined by a combination of heat, time and pH that can be optimized for each assay type. CleanAmp[™] dNTPs have moved to increase amplification specificiti and vield in a varieb or dNTPs have proved to increase amplification specificity and yield in a variety of

We will demonstrate the use of CleanAmp™ dNTPs in robust Multiplex Real-We will demonstrate the use of CleanAmp[™] dNPs in robust Multiplex Real-Time PCR and RFPCR assays that are designed for screening and detection of pathogenic bacteria and viruses in patient stool samples. Primer sets were designed to consensus regions of several strains of each pathogen to maximize detection. Assays were optimized using positive control baccterial isolates and were then tested on DNA and RNA isolated from patient stool samples that may contain inhibitors. Using commercially available CleanAmp[™] 2x RFPCR Master Mixis we aim to establish and standardize several constrained that detection and the optimation of the analysis. assay panels for detection of the pathogens causing infectious diarrhea that will make diagnosis and treatment faster and more reliable





Figure 4: One-Step RT-PCR Specificity is Improved by Hot Start Activation of Both the RT and PCR Steps



Figure 5: CleanAmp™ Enables Increased Accuracy and Consistent Results with Multiplexed One-Step RT-PCR





Figure 6: Standardized Real-Time Multiplex Assay Panels for Detection of Pathogenic Bacteria and Viruses

opment of standardized real-time multiplex assays: ner sets and hydrolysis probes were designed and tested for yield and specificity for

- individual target genes Optimal primers were combined for each assay and screened against well established
- ported particle received conservation of calcer values of the access degrates was conclusion accessed reference strating politive for specific participages [linical stool samples (previously validated) were screened for verification of positive arthogen detection

ogen detection says designed to include primers to detect an endogenous internal control (16S rRNA)



Figure 7: Real-Time Multiplex PCR Detection of Enterotoxigenic E. coli



Figure 8: Real-Time Multiplex PCR Detection of Various Types of Pathogenic E. coli



Figure 9: Pushing the Limit of Detection of E. coli Supplement Assay



Figure 10: RT-PCR Detection of Enteric Viruses in Patient Stool Samples



Conclusion

- CleanAmp^ $\ensuremath{^{\text{M}}}$ deprotection kinetics allow for application to a wide variety of biological assays.

- CleanAmp™ 2X Multiplex Master Mix allows for efficient amplification of multiple targets simultaneously
- CleanAmp[™] 2X RT-PCR Master Mix improves specificity by Hot Start control at both the RT and PCR steps.
- CleanAmp™ 2X RT-PCR Master Mix efficiently amplifies multiple targets at once.
- ETEC Toxins Assay and *E. coli* Supplement Assay successfully identify positive pathogens using CleanAmp[™] 2X Multiplex Master Mix. - E. coli Supplement Assay has a low limit of detection with CleanAmpTM 2X
- Multiplex Master Mix.
- CleanAmp^M 2X RFPCR Master Mix identifies positive pathogens by single-plex reactions for Enteric Viruses.

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The Modified Nucleic Acid Experts" www.trilinkbiotech.com