

Development of Standardized Pathogen Detection Assays Using CleanAmp™ Master Mixes

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Abstract

Acute gastroenteritis (AGE) affects over 179 million people and causes over 600,000 hospitalizations and 5000 deaths in the U.S. annually (CDC). Infectious AGE can be caused by a wide variety of different pathogens including bacteria, viruses and parasites. Rapid, definitive and economical identification of the causative AGE agent could inform medical decisions while reducing the inappropriate use of antibiotics. Standardized molecular methods and multiplexing will make pathogen identification more efficient and economical, with a huge potential global impact.

The molecular diagnostics field demands assays with more speed, specificity, and sensitivity. PCR is an established and reliable technology that meets these criteria. Although there are a myriad of published protocols using PCR for detection of specific diseases there is still a lack of established and standardized assays that cover the detection of certain infections.

CleanAmp™ dNTPs are a universal Hot Start technology that can be applied to many different types of PCR assays including real-time PCR, multiplex PCR and RT-PCR. CleanAmp™ dNTPs are 2'-deoxynucleoside triphosphates with a 3' thermolabile protecting group that prevents incorporation of the dNTPs at room temperature. As temperature is increased the protecting group begins to fall off creating a standard dNTP that can be naturally incorporated into the growing cDNA or PCR strand. CleanAmp™ dNTPs do not have a specific activation temperature but their kinetics are determined by a combination of heat, time and pH that can be optimized for each assay type. CleanAmp™ dNTPs have proved to increase amplification specificity and yield in a variety of assays.

We will demonstrate the use of CleanAmp™ dNTPs in robust multiplex real-time PCR and RT-PCR assays that are designed for screening and detection of pathogenic bacteria and viruses in patient stool samples. Primer sets were designed to consensus regions of several strains of each bacterial species to maximize detection. Assays were optimized using positive control bacterial isolates and then tested on DNA and RNA isolated from patient stool samples that may contain inhibitors. Using commercially available CleanAmp™ Master Mixes (CleanAmp™ 2x Multiplex Master Mix and CleanAmp™ 2x RT-PCR Master Mix) we aim to establish and standardize several assay panels for detection of AGE pathogens that will make diagnosis and treatment faster and more reliable.

Figure 1: CleanAmp™ Hot Start dNTP Structure & Activation

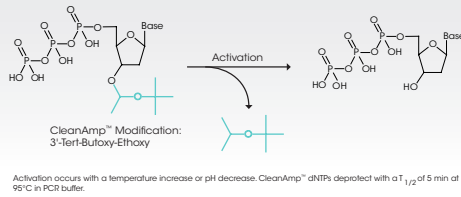


Figure 2: CleanAmp™ Deprotection Kinetics Allow for Application to a Wide Variety of Assays

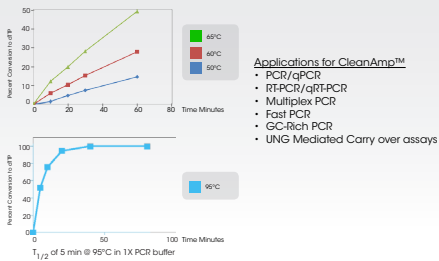


Figure 3: Efficient Fast Multiplex PCR with CleanAmp™ dNTPs

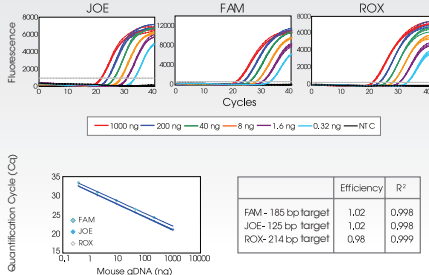


Figure 4: One-Step RT-PCR Specificity is Improved by Hot Start Activation of Both the RT and PCR Steps

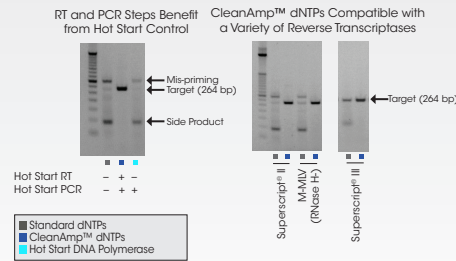


Figure 5: CleanAmp™ Enables Increased Accuracy and Consistent Results with Multiplexed One-Step RT-PCR

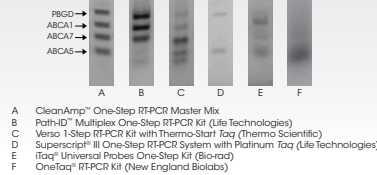


Figure 6: Standardized Real-Time Multiplex Assay Panels for Detection of Pathogenic Bacteria and Viruses

Development of standardized real-time multiplex assays:

- Primer sets and hydrolysis probes were designed and tested for yield and specificity for individual target genes
- Optimal primers were combined for each assay and screened against well established bacterial reference strains positive for specific pathogens
- Clinical stool samples (previously validated) were screened for verification of positive pathogen detection
- All assays designed to include primers to detect an endogenous internal control (16S rRNA)

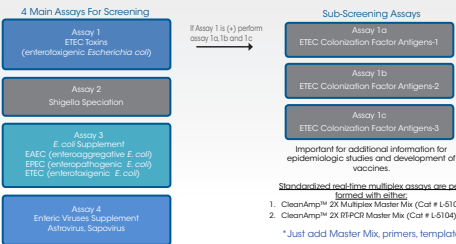


Figure 7: Real-Time Multiplex PCR Detection of Enterotoxigenic E. coli

Assay 1 - ETEC Toxins	Target Gene	Sample Type				NTC
		Bacterial Control Lysates	Stool Samples			
STe (sporadic heat stable enterotoxin)	estA1	+	+	+	+	
STh (human heat stable enterotoxin)	estA2-4	+		+	+	
LT (heat labile enterotoxin)	eitB1	+	+	+	+	
rmsA (16S rRNA)	rmsA	+	+	+	+	

+ Pathogen Detected.
Note: Multiple pathogens may contribute to infection and result in more than one positive per sample.

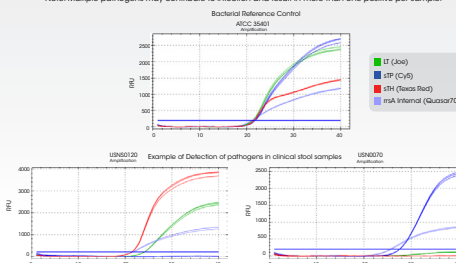


Figure 8: Real-Time Multiplex PCR Detection of Various Types of Pathogenic E. coli

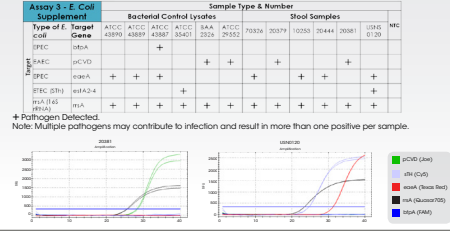


Figure 9: Pushing the Limit of Detection of E. coli Supplement Assay

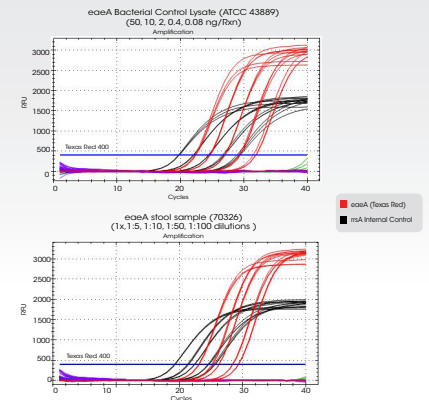
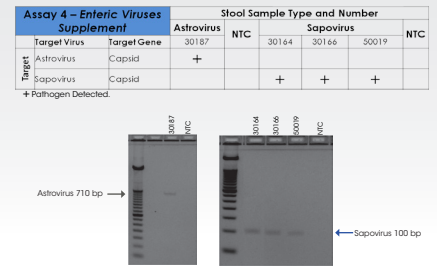


Figure 10: RT-PCR Detection of Enteric Viruses in Patient Stool Samples



Conclusion

- CleanAmp™ deprotection kinetics allow for application to a wide variety of biological assays.
- CleanAmp™ 2X Multiplex Master Mix allows for efficient amplification of multiple targets simultaneously.
- CleanAmp™ 2X RT-PCR Master Mix improves specificity by Hot Start control at both the RT and PCR steps.
- CleanAmp™ 2X RT-PCR Master Mix efficiently amplifies multiple targets at once.
- ETEC Toxins Assay and E. coli Supplement Assay successfully identify positive pathogens using CleanAmp™ 2X Multiplex Master Mix.
- E. coli Supplement Assay has a low limit of detection with CleanAmp™ 2X Multiplex Master Mix.
- CleanAmp™ 2X RT-PCR Master Mix identifies positive pathogens by single-plex reactions for Enteric Viruses.

Acknowledgments

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 Reaction conditions can be found at www.trilinkbiotech.com/posters
 Contact: Sabrina Shore, sshore@trilinkbiotech.com