

Description

Adenosine-5'-triphosphate (ATP), cytidine-5'-triphosphate (CTP), guanosine-5'-triphosphate (GTP), and uridine-5'-triphosphate (UTP) are unmodified ribonucleoside triphosphates, serving as essential building blocks of RNA molecules. Each of them consists of an unmodified base, a ribose, and a 5' triphosphate group. They can be used with modified nucleotides and TriLink's CleanCap® analogs in RNA synthesis by *in vitro* transcription.

TriLink offers both research-use-only (RUO) and good manufacturing practice (GMP) rNTPs. TriLink's GMP rNTPs are manufactured in highly controlled environments with documented procedures, traceability of materials and processes, and strict quality control measures, providing exceptional consistency, purity, and safety.

TriLink's RUO and GMP rNTPs may be ordered using the following catalog numbers:

| | RUO | | | | GMP | | |
|-----------------|-----------|-----------|------------|-----------|---------------|----------------|------------|
| | 10 µmol | 25 µmol | 100 µmol | bulk | 1 mmol | 10 mmol | bulk |
| ATP | N-1501-10 | N-1501-25 | N-1501-100 | N-1501-bk | FN-1501-1mmol | FN-1501-10mmol | FN-1501-bk |
| CTP | N-1502-10 | N-1502-25 | N-1502-100 | N-1502-bk | FN-1502-1mmol | FN-1502-10mmol | FN-1502-bk |
| GTP | N-1503-10 | N-1503-25 | N-1503-100 | N-1503-bk | FN-1503-1mmol | FN-1503-10mmol | FN-1503-bk |
| UTP | N-1504-10 | N-1504-25 | N-1504-100 | N-1504-bk | FN-1504-1mmol | FN-1504-10mmol | FN-1504-bk |
| rNTP set | N-1505-10 | N-1505-25 | N-1505-100 | N-1505-bk | — | — | — |

They are provided 100 mM in H₂O, pH 7.5.

Use & handling

Store at or below -20°C. Upon first use, prepare aliquots to minimize freeze-thaw cycles. Use only certified RNase-free reagents and consumables with proper RNase-free technique.

QC analysis

RUO products

- pH
- Concentration: UV/Vis
- Purity: ³¹P NMR, AX-HPLC
- Identity: ¹H NMR, mass spectrometry

GMP products (in addition RUO analyses)

- Characterization: appearance, residual chemicals
- Safety: endotoxin, bioburden
- Nuclease contamination: DNase and RNase detection

Product released by Quality Assurance. TriLink is certified ISO 9001:2015.

Troubleshooting

For any questions or technical support around this product, please reach out to support@trilinkbiotech.com

Other products in this guide

CleanCap Reagent AU is designed for the co-transcriptional capping of self-replicating or self-amplifying RNAs (saRNAs) with >95% capping efficiency¹. saRNA constructs are based on the genomes of positive sense (+) strand RNA viruses such as Venezuelan equine encephalitis virus (VEEV), Semliki forest virus (SFV), and Sindbis virus (SIN). The (+) strand genomes of these viruses start with a 5'-AU... Cap-1 saRNAs produced by CleanCap AU are more suitable for eukaryotic usage than Cap-0 saRNAs produced by legacy co-transcriptional capping methods such as anti-reverse cap analog (ARCA).

Modified uridines such as N1-methylpseudouridine, 5-methoxyuridine, and pseudouridine can reduce immunogenic response and enhance translational efficiency of saRNAs. These properties can result in safer saRNA and increased protein expression.

Related TriLink products

Adenosine-5'-Triphosphate, ATP (cat no. N-1501)
Cytidine-5'-Triphosphate, CTP (cat no. N-1502)
Guanosine-5'-Triphosphate, GTP (cat no. N-1503)
Uridine-5'-Triphosphate, UTP (cat no. N-1504)
rNTP Set: ATP, GTP, CTP, UTP (cat no. N-1505)
rNTP Set: ATP, CTP, GTP, N1MePsUTP (cat no. N-1506)*
rNTP Set: ATP, CTP, GTP, 5moUTP (cat no. N-1507)
rNTP Set: ATP, CTP, GTP, PsUTP (cat no. N-1508)

CleanCap[®] Reagent M6 (cat no. N-7453)
CleanCap[®] Reagent AG (cat no. N-7113)
CleanCap[®] Reagent AG (3' OMe) (cat no. N-7413)
CleanCap[®] Reagent AU (cat no. N-7114)

N1-Methylpseudouridine-5'-Triphosphate (cat no. N-1081)*
5-Methoxyuridine-5'-Triphosphate (cat no. N-1093)
Pseudouridine-5'-Triphosphate (cat no. N-1019)

T7 RNA Polymerase (Alphazyme cat no. E057)
Inorganic Pyrophosphatase (E. coli) (Alphazyme cat no. E051)
Engineered RNase Inhibitor (Alphazyme cat no. E07)

CleanCap[®] M6 EGFP mRNA (N1MePsU) (cat no. L-8101)[‡]
CleanCap[®] M6 FLuc mRNA (N1MePsU) (cat no. L-8102)
CleanCap[®] M6 mCherry mRNA (N1MePsU) (cat no. L-8103)
CleanCap[®] M6 Cas9 mRNA (N1MePsU) (cat no. L-8106)[§]
CleanCap[®] M6 EPO mRNA (N1MePsU) (cat no. L-8109)

Related TriLink services

TriLink offers RUO and GMP custom mRNA production services in addition to our catalog offerings. Visit our website trilinkbiotech.com/mrna-cdmo-services or contact us at mrna-services@trilinkbiotech.com for more information.

¹Final capping is dependent upon the CleanCap[®] Reagent, DNA template, and final mRNA sequence. Secondary structure due to RNA length and base composition can affect final capping efficiency, mRNA yield, and translation efficiency.

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Quick start protocol for *in vitro* transcription (IVT) with CleanCap AU

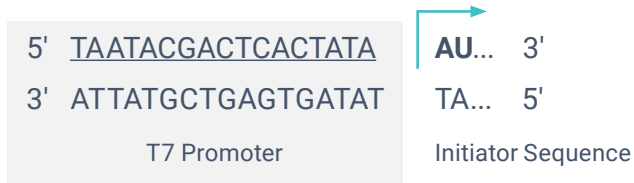
trilinkbiotech.com/cleancap-reagent-au.html

RNase-free techniques

It is essential that all reagents be RNase-free. Use disposable RNase-free tubes and bottles. Surfaces and pipettes can be wiped down with RNaseZap to destroy RNases. When possible, use dedicated RNase-free pipettes. Avoid using pipettes that have been used for plasmid preparation using RNase A.

Template design

Template design is an integral part of any transcription. CleanCap AU is to be used with the initiating sequence 5' AU 3'. The figure below shows the correct T7 promoter sequence (underlined) and initiator sequence (bold) for CleanCap AU.



Customer supplied materials

NOTE: All reagents must be RNase-free. Use recommended source or equivalent grade.

Required reagents

- DNA template
- Nucleoside-5'-Triphosphate (NTP) Set (TriLink cat no. N-1505)
Also available individually for use with modified NTPs. See related products for commonly used modified NTPs.
- T7 RNA polymerase (New England BioLabs cat no. M0251S)
- Inorganic Pyrophosphatase (New England BioLabs cat no. M0361S or M2403S)
- Murine RNase Inhibitor (New England BioLabs cat no. M0314S)
- 1 M Tris-HCL (pH 8.0), RNase Free (Thermo Fisher Scientific cat no. AM9856)
- Dithiothreitol (DTT) (EMD Millipore cat no. 3860-5GM)
- Spermidine (Sigma Aldrich cat no. 85558-1G)
- Triton X-100 (VWR cat no. 80503-490)
- 1 M Magnesium Acetate (Sigma Aldrich cat no. 63052)
- UltraPure™ DNase/RNase-Free Distilled Water (Thermo Fisher Scientific cat no. 10977015)

Optional reagents

- RNaseZap™ RNase Decontamination Solution (Thermo Fisher Scientific cat no. AM9780)

Reagent preparation and protocol

Reagent preparation

Add reagents in the prescribed order to ensure efficient transcription and capping. Ensure each component is homogenous before use. Store thawed enzymes on ice. These reaction conditions have been tested with templates up to 6 kb in length.

RNase-free techniques

It is essential that all reagents be rigorously RNase free. Use disposable RNase-free tubes and bottles. Surfaces and pipettes can be wiped down with RNaseZap to destroy RNases. When possible, use dedicated RNase-free pipettes. Avoid using pipettes that have been used for plasmid preparation using RNase A.

10X Transcription Buffer

400 mM Tris-HCL (pH 8)
100 mM DTT
20 mM Spermidine
0.02% Triton
270 mM Magnesium Acetate
DNase/RNase-Free Water

Transcription reaction

Add reagents in the prescribed order to ensure efficient transcription and capping. Store thawed enzymes on ice.

1. Add RNase-free water and NTPs to reaction tube.
2. Add CleanCap AU to tube and vortex to mix. Spin briefly to collect liquid.
3. Add 10X Transcription Buffer. Vortex. Spin briefly to collect Liquid.
4. Add DNA template.
5. Add Murine RNase Inhibitor, Yeast Inorganic Pyrophosphatase, and T7 RNA Polymerase.
6. Mix well by flicking or inverting tube 10 times and spin briefly to collect liquid.
7. Incubate at 37°C for 2-3 hours.

Table 1: Reaction components

| Component | Final | 100 μ L rxn |
|--|----------------------------------|---------------------------------------|
| DNase/RNase-free water | Up to 100 μ L | Up to 100 μ L |
| ATP (100 mM) | 5 mM | 5 μ L |
| CTP ¹ (100 mM) | 5 mM | 5 μ L |
| GTP (100 mM) | 5 mM | 5 μ L |
| UTP ¹ (100 mM) | 5 mM | 5 μ L |
| CleanCap AU (100 mM) | 4 mM | 4 μ L |
| 10X Transcription Buffer | 1X | 10 μ L |
| DNA template | 50 or 25 μ g/mL ² | 5 μ g or 2.5 μ g ² |
| Murine RNase Inhibitor (40 units/ μ L) | 1 unit/ μ L | 2.5 μ L |
| Yeast Inorganic Pyrophosphatase (0.1 units/ μ L) | 0.002 units/ μ L | 2 μ L |
| T7 RNA Polymerase (50 units/ μ L) | 8 units/ μ L | 16 μ L |
| Total Volume | 100 μL | 100 μL |

¹ Modified NTP can be used in place of wild-type. If using a modified NTP, use at the same concentration as the replaced wild-type NTP.

² Final concentration of DNA template should be 50 μ g/mL for a plasmid template or 25 μ g/mL for a PCR template.

CleanCap® products | Research license agreement

trilinkbiotech.com/cleancap-research-license

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